REVIEW

Mechanistic relationship between obesity-induced inflammation triggering endothelial dysfunction and the initiation of atherosclerosis development

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Abstract

Obesity is a multifactorial, chronic inflammatory illness that affects individuals of all ages; it is linked to several cardiovascular and metabolic syndromes. Excessive accumulation of fat that impairs metabolic processes is a key feature of obesity. Globally, more than 1.9 billion adults are overweight or obese, and the number of people affected by obesity is increasing. Obesity contributes to cardiovascular disease (CVD) and metabolic disease (MD) through processes such as inflammation, hyperglycemia, and dyslipidemia, as well as metabolic syndrome and obstructive sleep apnea. Furthermore, alterations in adipose tissue composition in obesity play a major role in the production of inflammatory cytokines that promote endothelial dysfunction (ED). Endothelial dysfunction is the first step in the mechanisms underlying obesity-related complications, such as atherosclerosis, which is a major complication of obesity. A growing body of evidence suggests that obesity-related ED makes a significant contribution to the development of CVD and MD.

Several mechanisms explain the association between obesity and atherosclerosis. The aim of this review is to summarize the effects of obesity-related inflammation on ED and its progression to atherosclerosis, focusing on cellular senescence, vascular aging, epigenetic modifications, reactive oxygen species, vascular calcification, and gut microbiota. In addition, we are also exploring new therapeutic strategies to reverse ED to prevent CVD and MD. Therefore, understanding the mechanisms underlying obesity-induced ED and its effects on atherosclerosis is crucial for developing therapeutic interventions.

Key words: Obesity, inflammation, reactive oxygen species, adipose tissue, endothelial dysfunction, atherosclerosis.

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Introduction

Obesity is a major public health issue worldwide. Excessive body fat storage without energy conversion plays a major role in obesity. Healthcare expenses, morbidity, and mortality are all substantially affected by obesity. It has reached epidemic proportions in many countries, affecting not only adults but also children and adolescents (1).

Cardiovascular (CV) disease (CVD) is the leading cause of death worldwide. Obesity is the most important risk factor for the development of coronary artery disease (CAD) (2). Some high-income nations have already experienced an initial obesity pandemic wave, whereas some low- and middle-income

nations have experienced a second wave. More than 25% of adults in some middle-income countries, such as South Africa, Libya, Egypt, Mexico, and Argentina, are reportedly afflicted with obesity. Compared to the rate of obesity prevalence just 20 years ago, this is a significant increase (3).

Obesity increases the risks of atherosclerosis (AS), stroke, type 2 diabetes mellitus (T2DM), and cardiovascular events (Fig. 1) (4, 5). According to the Global Burden of Disease survey reported in 2020, 4 million deaths, of which 2.7 million were CV, were attributable to high body mass index (BMI) (6). Obesity has attracted considerable attention as a global public health concern.

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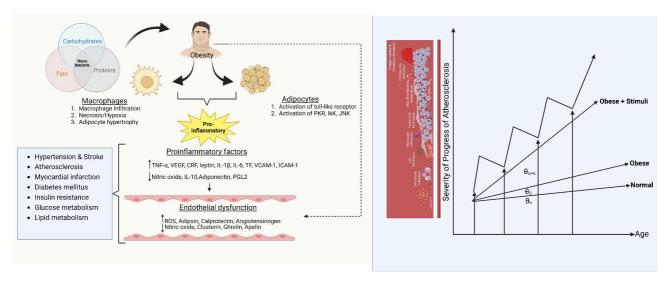
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Graphical abstract



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One type of obesity, known as metabolically healthy obesity (MHO), lacks cardiometabolic risk factors, such as hypertension, T2DM, insulin resistance and dyslipidemia (7).

Individuals with the MHO phenotype do not seem to develop AS (8).

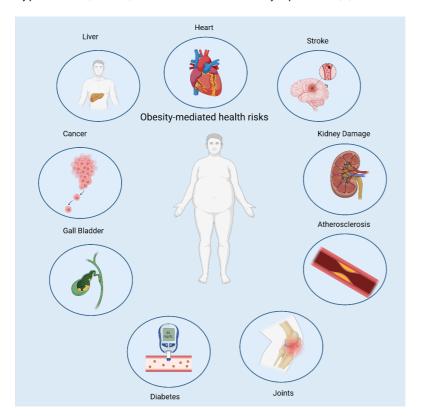


Figure 1. Comorbidities associated with obesity

Although various factors such as oxidative stress, loss of autophagic flux, macrophage NOD-like receptor protein 3 (NLRP3), inflammasome activation, adipokine imbalance, altered gut microbiome, and vascular inflammation are involved in obesity-induced AS, endothelial dysfunction (ED) plays a major role in CVD progression. In addition to genetic and environmental factors, obesity has been suggested to play a major role in the development of ED and AS (9).

Physiologically, vasoconstricting and vasodilating factors are secreted and released from ECs in a balanced manner (Fig. 2). Obesity-mediated AS disrupts this balance, thereby promoting the development of vascular ED and organ

damage (10). Atherosclerosis commences in childhood when cholesterol esters from macrophage foam cells are ingested, and the deposition of these esters on the wall surfaces of blood vessels leads to arterial wall thickening. Eventually, fatty streaks are formed owing to further lipid accumulation, which appears in young adults (11, 12).

Recently, much attention has been paid to the mechanisms underlying obesity-mediated ED, which have substantial implications for the prevention and treatment of CVD. The current review focuses on elucidating the pathophysiological mechanisms by which obesity-induced ED leads to AS and identifying therapeutic targets for the treatment of obesity.

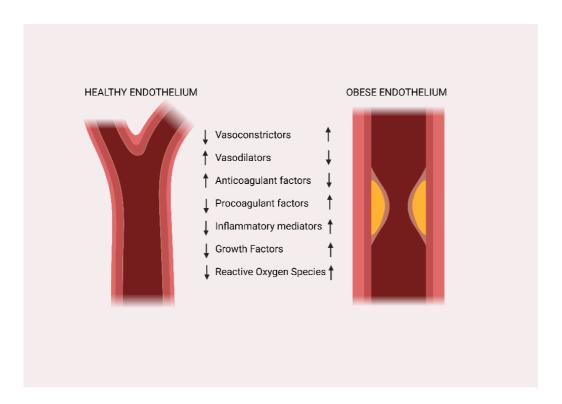


Figure 2. Differences in function of endothelial cells in healthy patients and patients with obesity

An overview of the mechanisms and their importance

Signaling mechanisms that contribute to ED in obesity-induced AS include autocrine, paracrine, and endocrine signaling. Studies have shown that tumor necrosis factor alpha (TNF- α) induces reactive oxygen species (ROS) production in small visceral fat arteries from patients with obesity, reducing nitric oxide (NO) availability (13).

Using in vivo and ex vivo myographic techniques, a reduction in endothelial function in the vascular region has been demonstrated, and the potential mechanisms include ROS-mediated rather than endothelial nitric oxide synthase (eNOS)-mediated dysfunction (13).

Classically, proinflammatory cytokines released by adipocytes are believed to play a major role in ED. Major adipokines generated by fat cells, such as resistin, leptin, and adiponectin,

play important roles in inflammation. Leptin, one of the crucial proteins produced by adipocytes, enhances the secretion of interleukin-6 (IL-6); furthermore, TNF-α directly or indirectly increases the levels of ROS, resulting in ED (14). Additionally, obesity damages the endothelial glycocalyx, which regulates NO production by responding to blood vessel forces. It was found that by thinning of the glycocalyx in mice and humans because of obesity, flow-mediated vasodilation was prevented in mesenteric arteries (15). Hindrance in endothelial properties causes the vascular endothelium to adopt a proinflammatory, prothrombotic, and proatherogenic phenotype, resulting in the pro-oxidation of mitogens, leukocyte adhesion, and platelet activation, as well as an increase in the levels of vasoconstriction factors, decreased synthesis of endothelium-derived hyperpolarizing factors (EDHFs), and impaired endothelial NO production (16) (Fig. 3).

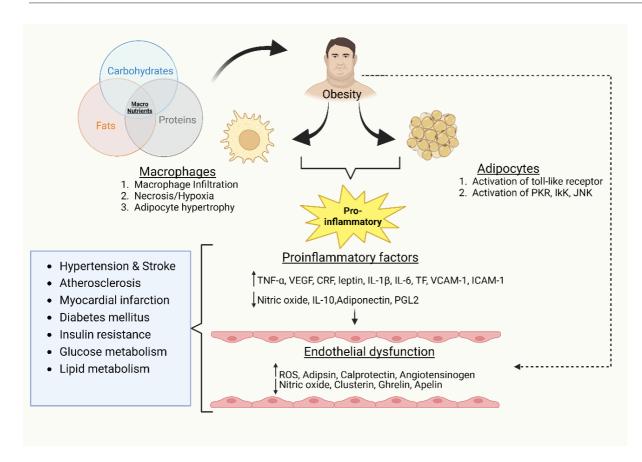


Figure 3. Pathological molecular mechanisms of obesity-induced endothelial dysfunction contributing to other diseases

Endothelial dysfunction causes CVD progression in obesity by activating adhesion molecules, leukocyte proliferation, and transmigration (17). Furthermore, dysfunctional adipocytes secrete angiotensinogen from the renin-angiotensin system (RAS), which causes its overexpression in RAS, enhanced ROS production, and atherogenic and thromboembolic potential of ECs (18). Adipose tissue is typically composed primarily of adipocytes and pre-adipocytes, with only minimal leukocytes. However, during obesity, the configuration and function of adipose tissue are largely altered. Endothelial cells secrete endocan (also known as endothelial cell-specific molecule-1), a soluble proteoglycan. The expression of endocan can be upregulated by inflammatory factors like TNF-α and IL-1β, which can then influence the expression of cell-adhesion molecules like vascular cell adhesion molecule-1 (VCAM-1) and intercellular adhesion molecule-1 (ICAM-1). These molecules are crucial for promoting leukocyte migration and the inflammatory response. Elevated plasma endocan levels are considered a possible inflammatory marker associated with CVD, which reflect endothelial activation and dysfunction (19).

Furthermore, endoglin, also known as CD105 or transforming growth factor beta receptor-3 (TGF- β receptor III), a homodimer transmembrane glycoprotein, is one of the ED inducers. Under several circumstances involving endothelial damage,

activation, inflammation, and endothelial senescence, soluble endoglins enter the systemic circulation. Furthermore, their levels increase during the early stages of AS and decreased in the later stages (20). Consequently, endoglin and endocan have been studied as potential biomarkers of ED and AS.

Adipose tissue alteration during obesity

White and brown adipose tissue are typical of mammals. Most of the mammalian adipose tissue, which is labeled as the area of energy storage, is white (21). White adipose tissue acts as a layer of thermal insulation, energy reservoir, mechanical protection tissue, and endocrine organs. Brown adipose tissue serves as an energy combustion site that maintains a stable body temperature. White adipose tissue accounts for 30-40% of the total body weight in lean women and 15-25% in lean men, whereas brown adipose tissue accounts for only 1–2% of fat in adult humans (22). In newborns, brown adipose tissue is typically elevated and works well in controlling energy expenditure and body temperature during non-shivering thermogenesis induced by uncoupling protein-1 (UCP1) (23). Visceral adipose tissue secretes more adipokines than subcutaneous adipose tissue, and these adipokines modulate adipocyte differentiation, lipid and glucose metabolism, neuroendocrine and cardiovascular functions, inflammatory processes and immunity (24).

Adipokines, chemokines, and cytokines, which are primarily elevated in obesity, are among the inflammatory factors produced by the adipose tissue, and an imbalance between pro- and anti-inflammatory factors plays a major role in obesity-mediated AS (25) (Fig. 4).

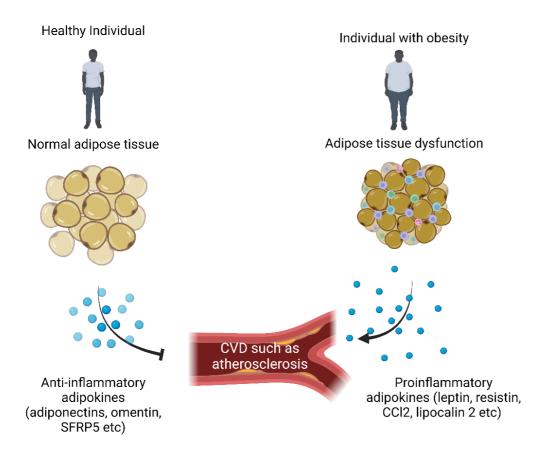


Figure 4. Physiological and pathological role of adipose tissue contributing to atherosclerosis

In addition, a different subset of UCP1-positive cells, called beige adipocytes, is prevalent in white adipocytes. Under the activation of sympathetic adrenergic receptors or cold stimulation, white adipocytes differentiate into brownlike adipocytes through a phenomenon called browning. Experimental reports have demonstrated the antiobesity and antidiabetic properties of beige adipocytes; consequently, they have garnered much attention for their therapeutic potential (26).

Adipose tissue inflammation is typically characterized by the infiltration of immune cells, including B and T lymphocytes, neutrophils, and macrophages. The primary cause of inflammation linked to ED is macrophage infiltration of adipose tissues.

Macrophage-specific genes, such as monocyte chemoattractant protein-1 (MCP-1), macrophage inflammatory protein-1 (MIP-1), F4/80, CD11b, and CD68, have been found to be enhanced in the obese mice adipose tissues.

Adipocyte size, F4/80 expression, and body mass were significantly correlated in adipose tissues after immunohistopathological analysis (27).

The composition, size, and structure of adipose tissue play a major role in the pathogenesis of obesity-induced CVD, such as AS. Adiposopathy or sick fat, a recently coined term for qualitative fat abnormalities, varies depending on an individual's lifestyle and physical activity (28). Clinical studies using cross-sectional abdominal imaging have shown that visceral adiposity has a stronger relationship with the incidence of systemic cardiometabolic rather than with subcutaneous fat quality (29). Although visceral fat contributes to only 5–20% of the total body fat mass compared with subcutaneous fat (≈ 80%), it contains an abundant source of IL-6, TNF-α, c-reactive protein in patients with abdominal obesity than in those with peripheral obesity (30). In addition, MRI or CT studies of the fat compartment demonstrated that systemic ED is significantly associated with visceral fat volume compared to subcutaneous fat volume, and gene-expression studies of visceral fat showed enhanced expression of antiangiogenic, proinflammatory, and oxidative stress-related genes (31-33).

Endothelial structure and function

The EC monolayer that lines the lumen of all blood vessels is known as endothelium. These cells serve as biocompatible barriers between all tissues and the flowing blood, which are both protective and insulating. The layers are arranged as follows: tunica intima, made up of ECs; the tunica media, which forms vascular smooth muscle cells (VSMCs); and tunica adventitia, which is an elastic lamina comprising

terminal nerve fibers around connective tissues (Fig. 5). The endothelial glycocalyx is the apical surface of the endothelium and is composed of glycoproteins, proteoglycans, and glycosaminoglycan chains (34, 35). Together, endothelial glycocalyx, secreted proteoglycans, and adsorbed plasma proteins form the endothelial surface layer. The ECs play a crucial role in homeostasis and arterial smooth muscle relaxation, and, consequently, vascular tone maintenance (36).

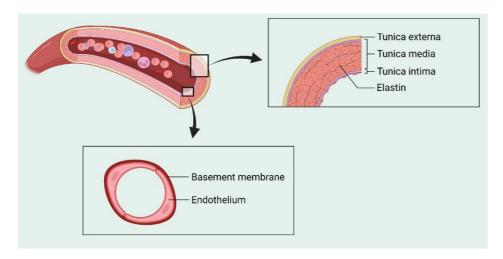


Figure 5. Structure of endothelium

The strategic location of endothelium helps to 'sense' changes in hemodynamic forces and blood-borne signals and 'respond' by releasing various autocrine and paracrine substances. Under normal physiological conditions, the balanced release of these bioactive compounds helps in maintaining vascular homeostasis (Fig. 6). Endothelium plays a major role in the

modulation of thrombogenicity, angiogenesis, vascular tone, inflammation, and dynamic permeability through receptor-mediated mechanisms and the balance between the release of autacoids, such as prostacyclin, endothelin-1, and angiotensin II (37-39).

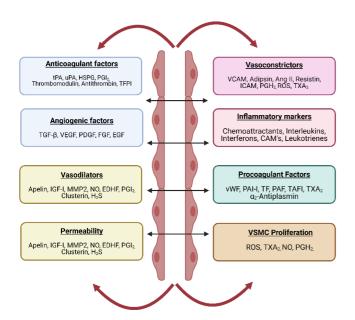


Figure 6. Important function of endothelial cells in regulating vascular homeostasis

The vascular endothelium produces prostacyclin (PGI2) and NO upon exposure to ecto-adenosine diphosphate (ADP)ase, CD73, and CD39, and prostaglandin E2 is promoted by glycosaminoglycans on the surface of ECs, thereby preventing platelet adhesion and aggregation. The endothelium also interacts with antithrombin III (ATH III), tissue factor pathway inhibitors (TFPIs), thrombomodulin (TM), and tissue factor (TF) to control adhesion and coagulation factor activity. In addition, the endothelium regulates urokinase plasminogen activator-1 (uPA-1), tissue-type plasminogen activator-1 (tPA-1), and inhibitors of uPA-1 and PAI-1. These mechanisms are influenced by the secretion of regulatory mediators, such as NO, endothelin-1 (ET-1), t-PA, PAI-1, prostanoids, adhesion molecules, angiotensin II (Ang II), von Willebrand factor (vWF), and cytokines. Endothelial cells secrete bioactive substances that control and maintain the structures and functions of intact blood vessels by balancing oxidative and antioxidative agents, VSMC proliferative and antiproliferative agents, inflammatory and anti-inflammatory factors, the blood coagulation cascade, vascular dilations and contractions, and the fibrinolytic system (40).

The endothelium exhibits anticoagulant activity via the synthesis of heparin sulfate proteoglycan, TM, and secretion of TFPIs. Further binding of TM to thrombin stimulates protein C, which inhibits factor VIII and factor V blood clotting. In addition, heparin-like molecules act as cofactors for ATH III, an anticoagulant molecule, whereas the complex activities of Factor Xa and TF-Factor VIIa are hindered by TFPIs.

Endothelial tissue plasminogen activator initiates fibrinolytic mechanisms, whereas NO and prostacyclin (PG12) are produced to control endothelial antiplatelet activities (41). The calcium–calmodulin complex activates eNOS to produce NO, L-arginine, nicotinamide adenine dinucleotide phosphate (NADPH), and tetrahydrobiopterin (BH4) as substrates. Furthermore, suppression of vasoconstriction and initiation of vasodilation are achieved by the absorption of NO by VSMCs, which then form a complex with the heteroiron group of guanylate cyclase. This complex, in turn, forms cyclic guanosine monophosphate (cGMP), activates cGMP-dependent protein kinase, and elevates Ca2+ levels in the cytosol within VSMCs.

Nitric oxide is a potent endothelium-derived vasodilator that possesses antiproliferative, antiplatelet, anti-inflammatory, and permeability-reducing properties. In addition, NO reacts with Kruppel-like factor 2 and EDHFs to enhance arterial vasodilation. Conversely, NO also inhibits MCP-1 and VCAM-1, thereby downregulating nuclear factor kappa B (NF-κB) expression (21).

Influence of endothelial dysfunction on cellular senescence and vascular aging

Atherosclerosis is an important cause of death in the Western world, which is associated with smoking, dyslipidemia, and hypertension as risk factors (48, 49). In addition to

disease-related ED, age-related ED plays an important role in CVD owing to its proinflammatory, vasoconstrictive, and prothrombotic environment(42), thereby disrupting vascular homeostasis.

Senescent cells are distinct in that they eventually cease to proliferate but do not eventually die, as they occur in a normal physiological process. Instead, the cells release substances that cause inflammation. Endothelial senescence is triggered by different senescence stressors, such as DNA damage (43), oxidative stress (44), oncogenic activation, and mitochondrial dysfunction (45).

Aged ECs show increased endothelin-1 (ET-1) expression, inflammation, and apoptosis, with attenuated endothelial NO production.

This results in the impairment of vessel tone, vascular integrity, and angiogenesis, resulting in the progression of CVD (46).

Adenosine monophosphate-activated kinase (AMPK), a mammalian target of rapamycin (mTOR), is a critical sensor of cellular homeostasis that influences endothelial senescence (47). A few clinical studies have shown that age-mediated arterial stiffness increases systolic pressure and decreases diastolic pressure, resulting in pulse pressure elevation and aortic pulse wave velocity (48).

Altered Ca2+ signaling in EC plays a major role in ED. Enhanced Ca2+ signaling promotes endothelium-dependent vasodilation via prostacyclin, NO, and EDHF. Physiologically, ECs play a major role in increasing Ca2+ levels and hyperpolarization by activating small-and intermediate-conductance Ca2+-activated potassium channels that initiate electrical conductance between vascular smooth muscle cells and ECs (46). However, all these pathways are impaired during aging, resulting in vascular dysfunction (Fig. 7).

Peroxynitrite, hydrogen peroxide, and hydroxyl radicals are the sources of ROS that contribute to age-related ED. In addition to ROS production through peroxisomal β-oxidation of fatty acid, mitochondrial respiration, lipoxygenase and xanthine oxidase, ROS levels are also increased via the uncoupling of eNOS, which results in the formation of peroxynitrite (49). Isoforms of nicotinamide adenine dinucleotide phosphate oxidase (NOX), such as NOX1, NOX2, NOX4, and NOX5, contribute to ED, apoptosis, and inflammation in AS (50). In addition, oxidative stress generates oxidized phospholipids (OxPLs) from polyunsaturated diacyl and alkaline (en) ylacyl glycerophospholipids.

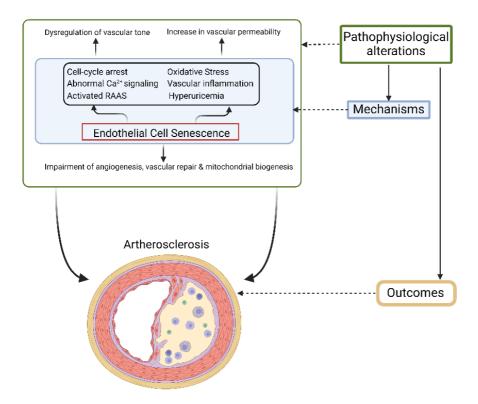


Figure 7. Pathophysiological mechanism of endothelial senescence resulting in atherosclerosis

Therefore, oxidative stress-induced cellular stress plays a major role in AS development. Various in vitro and in vivo studies have shown that OxPLs contribute to the development of AS. Briley et al. (51) demonstrated the presence of OxPLs in the atherosclerotic vessels of human lesions and in a hypercholesterolemia animal model. OxPLs contribute to the expression of proinflammatory cytokines in vascular ECs, thereby directly acting on leukocytes and influencing monocyte adhesion (52). Given their pertinent role in the development of numerous pathological conditions, OxPLs may function as biomarkers for the early detection of several CVDs, including AS.

Aging results in the enhanced expression of inflammatory cytokines, chemokines, and adhesion molecules in ECs. Interleukin-6, MCP-1, cellular adhesion molecules, IL-1 β and TNF- α are key cytokines that are overexpressed because of initiation of NF- κ B signaling resulting in endothelial dysfunction (50). In addition, activation of NLRP3 initiates oligomerization resulting in attracting procaspase-1, leading to production of ROS, chemokines and activation of NF- κ B (53). Inhibition of NF- κ B activation during prolonged TNF- α exposure prevents the premature senescence of ECs, thereby demonstrating a pathological association between inflammation and EC senescence (53, 54).

Oxidative stress and inflammation play major roles in the activation of the vascular renin-angiotensin-aldosterone system (RAAS). Atherogenesis is a critical process in

atherosclerotic plaque formation and is influenced by the activation of angiotensin II, which further decreases NO generation. Inflammation, altered lipid metabolism, and endothelial injury can lead to atherogenesis. Of these, endothelial injury is the initial trigger leading to the infiltration and accumulation of multiple modified low-density lipoprotein (LDL) particles in the endothelium (55). The accumulation of modified LDL initiates the production of VCAM-1 and P&E-selectins, resulting in the recruitment of leukocytes to the endothelial space, thereby attracting inflammatory cells, which are influenced by MCP-1.

Furthermore, macrophages express scavenger receptor class A (SRA), cluster of differentiation 36 (CD36), which act through modified LDL internalization, resulting in inflammation and ROS (56).

The senescence of ECs leads to CVD, and uric acid levels are higher in aged women than in men (57). Aortic stiffness is closely associated with the inflammatory response and maladaptive immunity. Inhibition of aortic xanthine oxidase activity (increased because of the western diet) by allopurinol decreases aortic stiffness and impairs endothelial-dependent arterial relaxation (58). A large multicenter study by Neogi et al. demonstrated an association between serum uric acid levels and carotid plaques (59). They also found this association in men, even in the absence of other CVD risk factors and coronary calcifications.

Epigenetic modifications

Histone acetylation and DNA methylation are examples of epigenetic modifications that are post-replication chromatin alterations that do not affect basal nucleotide code (60). These processes are linked to several age-related diseases, such as AS, and are crucial for regulating gene activation and silencing (61). Therefore, epigenetic changes have been used as biomarkers of cellular senescence in vascular ECs. Prior research has identified specific changes in methyltransferase expression, suggesting that these changes are associated with hypomethylation of hypermethylated genomic regions. These genes are involved in lipid metabolism, proliferation, and apoptosis. Atherogenic lipoproteins promote DNA methylation and histone deacetylation (60). These results suggest a possible association between AS and epigenetic alterations. Histone acetylation and deacetylation play major roles in the development of AS with age, and these changes can affect VSMC proliferation, extracellular matrix (ECM) composition, and inflammation. Moreover, additional research has demonstrated that the mitochondrial gene p66Shc, which is recognized as a gene associated with longevity, stimulates histone acetylation and hypermethylation, causing an agerelated increase in p66Shc synthesis and its consequent activation (62).

The Genetics of Healthy Aging Study demonstrated that genes encoding apolipoprotein E, forkhead box protein O1, IL-6, and Sirt6 are functionally associated with EC senescence (63). Among these, transcriptional expression and senescence were significantly correlated with argininosuccinate synthase 1, NOX4, aquaporin 1, p15, and p16. Telomere changes are a key mechanism in the senescence of vascular cells. Leukocyte telomere length is substantially correlated with vascular cell senescence, hypertension, and atherothrombotic events in a clinical study involving a cohort of 193 patients (≥70 years of age) (64). Therefore, telomerase activity controls both CVD and EC functions. Single-stranded RNAs of approximately 25 nucleotides, known as microRNAs (miRNAs), regulate the pathobiology of CVD and EC senescence. Overexpression of ASncmtRNA-2 induces replicative senescence in ECs and cellcycle arrest in the G2/M phase, indicating a potential role for these non-coding RNAs in the regulation of EC senescence, vascular function, and CVD (65).

Impact of oxidative stress

Oxidative stress markers are highly correlated with fat accumulation in humans with obesity (66). A higher risk of AS is associated with increased oxidant production on account of NO quenching. An imbalance between free radical generation and the antioxidant system can lead to oxidative stress. Increased production of ROS results from the reduced expression of antioxidative enzymes in adipose tissue, including glutathione peroxidase, catalase, and superoxide dismutase.

Oxidative stress linked to atherosclerotic risk factors such as obesity, can also trigger ED (67). According to the oxidative

theory of AS, oxidative alteration of lipoproteins causes lipid peroxidation, which, in turn, causes foam cell development and EC destruction in the arterial wall, both of which have a substantial impact on atherogenesis. Furthermore, isoprostanes (IsoPs) contribute to the etiology of vascular dysfunction through their activity. Strong vasoconstrictors in most arterial beds, such as IsoPs, also promote platelet aggregation and increase the adherence of neutrophils and monocytes to ECs, all of which may be involved in AS. Isoprostanes are prostaglandin-like compounds produced by free radical-induced peroxidation independent of cyclooxygenase. Furthermore, in individuals with obesity, oxidative stress is involved in various interactions with metabolism and atherogenic lipid activities. These interactions include a postprandial increase in the levels of triglycerides and oxidative modification of LDLs, resulting in the production of oxidized LDL (oxLDL), which has crucial atherogenic properties and a reduction in high-density lipoproteins. These interactions may result in increased ROS production in the endothelium, initiation of proinflammatory vascular processes, and increased endothelial damage, resulting in ED, all of which can lead to the initiation of atherosclerotic processes (68).

In addition to an increase in the levels of oxidative stress markers, such as thiobarbituric acid-reactive substances (TBARS) and protein carbonyls, non-high-density lipoprotein cholesterol (non-HDL-C) levels and the ratios of triglyceride/ high-density lipoprotein cholesterol (HDL-C), cholesterol/HDL-C, and LDL cholesterol (LDL-C)/HDL-C were also increased. In addition, the levels of antioxidant defense markers, such as glutathione (GSH), CAT, and GPx, were reduced (69). Furthermore, increased oxidized LDL levels have been linked to procoagulant and cytotoxic activities, foam cell production, growth, destabilization of atherosclerotic plagues, and increased expression of EC-adhesion molecules. Increased synthesis of vasoconstrictor proteins, oxidative stress, and impaired endothelial function reliant on NO are linked to an increased amount of total adipose tissue and degree of abdominal obesity. Patients who were overweight or affected by obesity had higher levels of phosphorylation of eNOS, catalase, nitrotyrosine, and NADPH oxidase-p47(phox) protein in vascular ECs than patients with normal weight.

A recent review highlighted the association between apolipoprotein M (ApoM), sphingosine-1-phosphate (S1P), HDL, and endothelial dysfunction. The HDL-bound apoM/S1P complex acts as a link between HDL and ECs under normal physiological conditions, preserving the integrity of the endothelial barrier. However, during pathological CV events, this apoM/S1P-signaling leads to inflammation-mediated endothelial dysfunction (70). This could offer a new perspective on the molecular processes that connect obesity to oxidative stress and a higher risk of atherosclerotic CVD (71). Similarly, López-Domènech et al. (72) demonstrated that weight loss results in a decrease in blood pressure, with a significant reduction in the levels of subclinical markers of AS,

such as myeloperoxidase (MPO), sP-selectin, small and dense LDL particles, and leukocyte adhesion.

Endothelial dysfunction and vascular calcification

Vascular calcification is involved in vascular remodeling owing to ED, dedifferentiation of VSMCs, and changes in elastin and collagen levels. Medial arterial, intimal, and infantile calcifications are the three primary forms of vascular calcification. Patients with T2DM, chronic renal failure, and advanced age frequently exhibit medial arterial calcification. Nonetheless, intimal calcification is thought to be a passive and degenerative process more commonly associated with AS. It is acknowledged as a self-regulating active process (73). The mechanism of vascular calcification is a tightly regulated, multifactorial process that resembles osteochondrogenesis. It has also been associated with dysregulated metabolism, inflammation, osteogenesis, and progressive AS. The aggressive inflammatory stage of AS is characterized by plaque calcification. Tumor necrosis factor-α and other inflammatory cytokines promote osteogenic differentiation and ECM mineralization. Matrix vesicles are produced by smooth muscle cells and macrophages and may act as sites of initiation for the development of mineral crystals, leading to microcalcifications. Microcalcifications grow into massive masses, spread into the surrounding collagenous matrix from the deeper part of the necrotic core, and ultimately coalesce into calcified sheets or plates (74).

In males, the visceral adiposity index is closely associated with the total coronary calcification score (75). Given that the coronary artery calcium score and total plaque burden are known to be highly correlated, this score can be used to assess the severity of coronary AS.

Impact of human gut microbiota

The gut microbiota (1014 bacteria) plays a vital role in maintaining immune system homeostasis and normal development, regulating the proliferation of epithelial cells, protecting against pathogenic bacteria, and influencing villus architecture and angiogenesis in the intestine (76). Research in both humans and mice have demonstrated that changes in the gut microbiota are linked to several illnesses, such as AS, diabetes mellitus, obesity, and inflammation. The first study in leptin-deficient (ob/ob) mice demonstrated the presence of more phylum Firmicutes and less phylum Bacteroidetes in obese ob/ob mice than in lean controls, and the microbiota have important roles in arterial blood pressure, which is again modulated by the endothelium. Further transfer of the microbiome from obese/lean mice to germ-free mice resulted in the corresponding phenotypes, thereby demonstrating the role of the microbiome in pathology (77, 78). Two main pathways exist by which intestinal bacteria can affect the endothelium of the circulatory system: first, the microbiota and its metabolites can activate the enteric nervous system, which in turn can affect the activity of brain centers that control the cardiovascular system; second, the bacteria can

enter the bloodstream through the blood-intestinal barrier and alter the function of the tissues and organs that regulate the circulatory system's homeostasis. The genus Collinsella is more prevalent in symptomatic atherosclerotic patients, whereas Roseburia and Eubacterium are more prevalent in healthy controls (79). Further metagenomic analysis showed that patient gut metagenomes were deprived of phytoene dehydrogenase and enhanced in genes encoding peptidoglycan synthesis; additionally, patients' β-carotene serum levels were also decreased. Through fermentation and decomposition, microbiota metabolize resistant starches and dietary fibers, and supply the host with short-chain fatty acids (SCFAs; (acetate, propionate, butyrate). Intestinal epithelial cells, immune cells, and adipocytes have various receptors, including G-protein coupled receptor 41/free fatty acid receptor 3 (GPR41/FFAR3), GPR43/FFAR2, GPR109A, and olfactory receptor 78, to which SFAs bind, despite their varying expression levels between tissues and cell types. Trimethylamine (TMA) and trimethylamine N-oxide (TMAO), metabolites derived from gut microbes, have been shown by Tang et al. to be proatherogenic in both humans and mice (80-82). Aguilar et al. (83, 84) demonstrated using ApoE knockout mice that a butyrate-supplemented chow diet decreased atherosclerotic lesions and further reduced the production of vascular cell adhesion molecule-1, chemotaxis protein-1 (CCL2/MCP-1), and matrix metalloproteinase-2 at the lesion site.

This resulted in increased collagen deposition, reduced macrophage migration, and increased plaque stability. In addition, macrophages from butyrate-treated mice exhibited low ROS and NO release.

The gut microbiota produces TMA, which enters the bloodstream and is oxidized to form TMAO, a cardiac risk biomarker in the liver. Trimethylamine N-oxide has been demonstrated to possess proatherogenic properties owing to ED and the prothrombotic effect caused by platelet aggregation. Activation of the transcription factor NF-kB, which mediates the upregulation of inflammatory signals and adhesion of leukocytes to ECs, results in TMAO-induced ED. Additionally, Seldin et al. demonstrated that CVD and high plasma levels of TMAO are linked to concurrent reduction in endothelial progenitor cells and increased inflammation (85). These studies provide strong evidence that the composition of the gut microbiota and their metabolites play a major role in ED and the resulting AS. Murphy et al. (86) demonstrated that acute infections and other transient stimuli are more likely to cause cumulative arterial damage leading to AS. Disease severity increases with the frequency of acute stimuli and can be made more severe by metabolic, environmental, and genetic risk factors. Animal studies have shown that transient vessel damage heals only partially when the infection is managed. However, repeated inoculation of mice with pathogens significantly enhanced atherosclerotic damage (87) (Fig. 8).

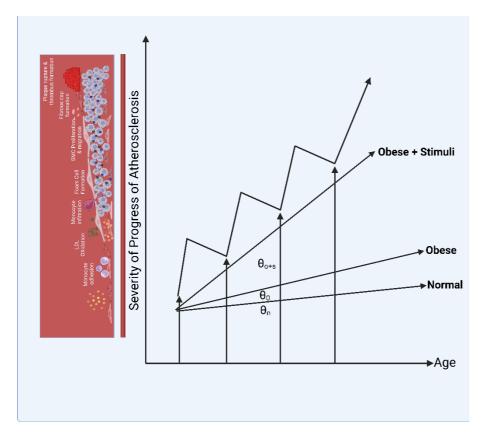


Figure 8. Atherosclerosis development during obesity because of cumulative repetitive infection and inflammatory insult

Reversal of endothelial dysfunction

The reversal of endothelial dysfunction is a promising strategy for preventing ED-mediated diseases. Antioxidants, vitamins, and pharmacological agents have been studied to understand the mechanisms of ED reversal properties (88). It may be possible to reduce the risk of acute CVD events by slowing the progression of AS, altering ED, or both. Free radicals play a major role in age-related endothelial impairment and showed that consumption of an antioxidant cocktail (Vitamin C, 1000 mg; Vitamin E, 600 I.U.; Alpha lipoic acid, 600 mg) restores endothelial function in the elderly, while impairing normal endothelium-dependent vasodilation in the young (89). In addition, various pharmacological agents, such as angiotensin-converting enzyme inhibitors, endothelium NO synthase enhancers, sphingosine-1-phosphate, statins, betablockers, phosphodiesterase 5 inhibitors, calcium-channel blockers, and angiotensin AT1 receptor blockers, have endothelial-protective effects (90).

Furthermore, despite their ability to improve glycemic control in humans and animal models of T2DM using dipeptidyl peptidase-4 (DPP4) inhibitors, they also protect against vascular aging through various intricate cellular mechanisms. These include enhancing ED, inhibiting the proliferation and migration of VSMCs, increasing the levels of circulating endothelial progenitor cells (EPCs), and hindering

the infiltration of mononuclear macrophages. In addition, DPP4 enhances EC proliferation and migration, mitigates senescence, and blocks apoptosis (91). Recently, the prevention of eNOS uncoupling has attracted considerable interest as a therapeutic option for preventing ED-mediated diseases. One of the most important chemicals secreted by EC is NO, the antiatherogenic properties of which promote cardiovascular homeostasis. Endothelial NO synthase generates vascular NO from the substrate I-arginine (I-Arg), with tetrahydrobiopterin (BH4) serving as the essential cofactor. Under pathological conditions, such as CVD, vascular oxidative stress hinders eNOS activity and favors it's uncoupling, leading to ED. Uncoupled eNOS generates superoxide anion (O2-) rather than NO, thereby serving as a source of dangerous free radicals that worsen oxidative stress. The key mechanisms underlying eNOS uncoupling include deficiency of the eNOS substrate L-arginine, oxidative depletion of the critical eNOS cofactor BH4, eNOS S-glutathionylation, and accumulation of its analog, asymmetrical dimethylarginine (ADMA). Hence, various pharmacological agents, such as sepiapterin (member of the pteridine class of organic chemicals), 5-methyltetrahydrofolate, fluvastatin, cerivastatin, L-arginine supplementation, and thioredoxin, are being studied to prevent eNOS uncoupling (92). Takase et al. (93) found that ezetimibe in combination with statins improved the functional prognosis and reduced ED in stented coronary arteries. It was also linked to greater reductions in oxysterol levels compared to statin monotherapy (93). Another studies demonstrated that the administration of statins along with eicosatetraenoic acid (EPA), an omega-3 polyunsaturated fatty acid, not only lowers plasma triglyceride levels but also has significant benefits against atherosclerotic plaques (94, 95). The potential benefits of EPA include lowering atherogenic parameters, such as non-HDL-C, oxidized LDL particles, triglycerides, and remnant-like particle cholesterol (RLP-C). It also increases the levels of antiatherogenic factors, thereby improving endothelial function, enhancing antioxidant properties, and secreting pro-resolving lipid mediators called resolvins (94, 96). Furthermore, they decrease macrophage accumulation, monocyte adhesion, and foam cell accumulation. These studies provide strong evidence that ED reversal holds promising therapeutic potential to prevent or at least delay AS.

Conclusion and future directions

Although it is difficult to isolate the specific mechanism underlying obesity-mediated AS, alterations in endothelial structure and function, cellular senescence, epigenetic factors, oxidative stress, vascular calcification, and gut microbiome play crucial roles in disease incidence and progression. Exploring the potential link between these factors not only helps to understand the mechanisms of the disease but also to find a multi-targeted therapeutic agent to prevent or slow down the AS process. Targeting these mechanisms through both prevention and treatment at an early stage can delay the progression of AS. Further lifestyle and diet changes are keys to prevent obesity in the first place.

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References

- Nagpal N, Messito MJ, Katzow M, Gross RS. Obesity in children. Pediatrics Rev 2022; 43: 601-17. doi: 10.1542/ pir.2021-005095
- Kozieł P, Jankowski P, Mirek-Bryniarska E, Nessler J, Podolec P, De Bacquer D, et al. Obesity in patients with established coronary artery disease over a 20-year period (1997-

- 2017). Polish Arch Int Med 2021; 131: 26-32. doi: 10.20452/pamw.15742
- Azab M, Al-Shudifat AE, Johannessen A, Al-Shdaifat A, Agraib LM, Tayyem RF. Are risk factors for coronary artery disease different in persons with and without obesity? Metab Syndr Rel Disord 2018; 16: 440-5. doi: 10.1089/ met.2017.0152
- Khan SS, Ning H, Wilkins JT, Allen N, Carnethon M, Berry JD, et al. Association of body mass index with lifetime risk of cardiovascular disease and compression of morbidity. JAMA Cardiol 2018; 3: 280-7. doi: 10.1001/ jamacardio.2018.0022
- 5. Kajikawa M, Maruhashi T, Kishimoto S, Hashimoto H, Takaeko Y, Yamaji T, et al. Association of body mass index with endothelial function in Asian men. Int J Cardiol 2021; 324: 186-92. doi: 10.1016/j.ijcard.2020.09.029
- Dai H, Alsalhe TA, Chalghaf N, Riccò M, Bragazzi NL, Wu J. The global burden of disease attributable to high body mass index in 195 countries and territories, 1990-2017: An analysis of the Global Burden of Disease Study. PLoS Med 2020; 17: e1003198. doi: 10.1371/journal.pmed.1003198.
- 7. Lassale C, Tzoulaki I, Moons KGM, Sweeting M, Boer J, Johnson L, et al. Separate and combined associations of obesity and metabolic health with coronary heart disease: a pan-European case-cohort analysis. Eur Heart J 2018; 39: 397-406. doi: 10.1093/eurheartj/ehx448.
- 8. Yoon JW, Jung CH, Kim MK, Park HE, Park KS, Jang HC, et al. Influence of the definition of "metabolically healthy obesity" on the progression of coronary artery calcification. PloS One 2017; 12: e0178741. doi: 10.1371/journal.pone.0178741
- Kajikawa M, Higashi Y. Obesity and endothelial function. Biomedicines 2022; 10. doi: 10.3390/ biomedicines10071745
- Sena CM, Pereira AM, Seiça R. Endothelial dysfunction

 a major mediator of diabetic vascular disease. Biochim
 Biophys Acta 2013; 1832: 2216-31. doi: 10.1016/j.
 bbadis.2013.08.006
- 11. McGill HC, Jr. Fatty streaks in the coronary arteries and aorta. Lab Investigat: J Tech Meth Pathol 1968; 18: 560-4.
- 12. Strong JP, Malcom GT, McMahan CA, Tracy RE, Newman WP, 3rd, Herderick EE, et al. Prevalence and extent of atherosclerosis in adolescents and young adults: implications for prevention from the Pathobiological Determinants of Atherosclerosis in Youth Study. JAMA 1999; 281: 727-35.. doi: 10.1001/jama.281.8.727
- 13. Virdis A, Santini F, Colucci R, Duranti E, Salvetti G, Rugani I, et al. Vascular generation of tumor necrosis factor-α reduces nitric oxide availability in small arteries from visceral fat of obese patients. J Am Coll Cardiol 2011; 58: 238-47. doi: 10.1016/j.jacc.2011.01.050

- Kershaw EE, Flier JS. Adipose tissue as an endocrine organ.
 J Clin Endocrinol Metab 2004; 89: 2548-56. doi: 10.1210/jc.2004-0395
- Fancher IS, Le Master E, Ahn SJ, Adamos C, Lee JC, Berdyshev E, et al. Impairment of flow-sensitive inwardly rectifying K(+) channels via disruption of glycocalyx mediates obesity-induced endothelial dysfunction. Arterioscl Thromb Vasc Biol 2020; 40: e240-e55. doi: 10.1161/atvbaha.120.314935
- 16. Xu J, Zou MH. Molecular insights and therapeutic targets for diabetic endothelial dysfunction. Circulation 2009; 120: 1266-86. doi: 10.1161/circulationaha.108.835223
- Barton M. Obesity and aging: determinants of endothelial cell dysfunction and atherosclerosis. Pflugers Arch: Eur J Physiol 2010; 460: 825-37. doi: 10.1007/s00424-010-0860-y
- Kwaifa IK, Bahari H, Yong YK, Noor SM. Endothelial dysfunction in obesity-induced inflammation: molecular mechanisms and clinical implications. Biomolecules 2020; 10. doi: 10.3390/biom10020291
- Chen J, Jiang L, Yu XH, Hu M, Zhang YK, Liu X, et al. Endocan: A key player of cardiovascular disease. Front Cardiovasc Med 2021; 8: 798699. doi: 10.3389/fcvm.2021.798699
- Li CG, Bethell H, Wilson PB, Bhatnagar D, Walker MG, Kumar S. The significance of CD105, TGFbeta and CD105/TGFbeta complexes in coronary artery disease. Atherosclerosis 2000; 152: 249-56. doi: 10.1016/s0021-9150(99)00476-1
- 21. Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. Nature Rev Immunol 2006; 6: 772-83. doi: 10.1038/nri1937
- 22. Cypess AM. Reassessing human adipose tissue. New Engl J Med 2022; 386: 768-79. doi: 10.1056/NEJMra2032804
- 23. Darvall KA, Sam RC, Silverman SH, Bradbury AW, Adam DJ. Obesity and Thrombosis. Eur J Vasc Endovasc Surg 2007; 33: 223-33. doi: 10.1016/j.ejvs.2006.10.006
- 24. O'Mara AE, Johnson JW, Linderman JD, Brychta RJ, McGehee S, Fletcher LA, et al. Chronic mirabegron treatment increases human brown fat, HDL cholesterol, and insulin sensitivity. J Clin Invest 2020; 130: 2209-19. doi: 10.1172/jci131126
- 25. Karastergiou K, Mohamed-Ali V. The autocrine and paracrine roles of adipokines. Mol Cell Endocrin 2010; 318: 69-78. doi: 10.1016/j.mce.2009.11.011
- 26. Yang X, Yi X, Zhang F, Li F, Lang L, Ling M, et al. Cytochrome P450 epoxygenase-derived EPA and DHA oxylipins 17,18-epoxyeicosatetraenoic acid and 19,20-epoxydocosapentaenoic acid promote BAT thermogenesis and WAT browning through the GPR120-AMPKα signaling pathway. Food Funct 2022; 13: 1232-45. doi: 10.1039/d1fo02608a

- Segovia SA, Vickers MH, Gray C, Reynolds CM. Maternal obesity, inflammation, and developmental programming. BioMed Res Int 2014; 2014: 418975. doi: 10.1155/2014/418975
- 28. Thompson D, Karpe F, Lafontan M, Frayn K. Physical activity and exercise in the regulation of human adipose tissue physiology. Physiol Rev 2012; 92: 157-91. doi: 10.1152/physrev.00012.2011
- 29. Rosenquist KJ, Pedley A, Massaro JM, Therkelsen KE, Murabito JM, Hoffmann U, et al. Visceral and subcutaneous fat quality and cardiometabolic risk. JACC Cardiovasc Imag 2013; 6: 762-71. doi: 10.1016/j.jcmg.2012.11.021
- Park HS, Park JY, Yu R. Relationship of obesity and visceral adiposity with serum concentrations of CRP, TNF-alpha and IL-6. Diabetes Res Clin Pract 2005; 69: 29-35. doi: 10.1016/j.diabres.2004.11.007
- 31. Gealekman O, Guseva N, Hartigan C, Apotheker S, Gorgoglione M, Gurav K, et al. Depot-specific differences and insufficient subcutaneous adipose tissue angiogenesis in human obesity. Circulation 2011; 123: 186-94. doi: 10.1161/circulationaha.110.970145
- 32. Alvehus M, Burén J, Sjöström M, Goedecke J, Olsson T. The human visceral fat depot has a unique inflammatory profile. Obesity 2010; 18:879-83. doi:10.1038/oby.2010.22
- 33. Ledoux S, Queguiner I, Msika S, Calderari S, Rufat P, Gasc JM, et al. Angiogenesis associated with visceral and subcutaneous adipose tissue in severe human obesity. Diabetes 2008; 57: 3247-57. doi: 10.2337/db07-1812
- 34. Jaffe EA. Cell biology of endothelial cells. Human Pathol 1987; 18: 234-9. doi: 10.1016/s0046-8177(87)80005-9
- 35. Luft JH. Fine structures of capillary and endocapillary layer as revealed by ruthenium red. Federation proceedings. 1966;25(6):1773-83. Epub 1966/11/01. PubMed PMID: 5927412.
- 36. Furchgott RF, Zawadzki JV. The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. Nature. 1980;288(5789):373-6. Epub 1980/11/27. doi: 10.1038/288373a0. PubMed PMID: 6253831.
- 37. Goligorsky MS. Endothelial cell dysfunction: can't live with it, how to live without it. Am J Physiol Renal Physiol 2005; 288: F871-80. doi: 10.1152/ajprenal.00333.20047
- Félétou M, Vanhoutte PM. Endothelial dysfunction: a multifaceted disorder (The Wiggers Award Lecture). Am J Physiol Heart Circ Physiol 2006; 291: H985-1002. doi: 10.1152/ajpheart.00292.2006
- 39. Reitsma S, Slaaf DW, Vink H, van Zandvoort MA, oude Egbrink MG. The endothelial glycocalyx: composition, functions, and visualization. Pflugers Arch: Eur J Physiol 2007; 454: 345-59. doi: 10.1007/s00424-007-0212-8

- 40. Mussbacher M, Salzmann M, Brostjan C, Hoesel B, Schoergenhofer C, Datler H, et al. Cell type-specific roles of NF-KB linking inflammation and thrombosis. Front Immunol 2019; 10: 85. doi: 10.3389/fimmu.2019.00085
- Katakami N. Mechanism of development of atherosclerosis and cardiovascular disease in diabetes mellitus. J Atheroscler Thromb 2018; 25: 27-39. doi: 10.5551/jat. RV17014
- 42. Donato AJ, Machin DR, Lesniewski LA. Mechanisms of dysfunction in the aging vasculature and role in agerelated disease. Circ Res 2018; 123: 825-48. doi: 10.1161/circresaha.118.312563
- 43. Zhan H, Suzuki T, Aizawa K, Miyagawa K, Nagai R. Ataxia telangiectasia mutated (ATM)-mediated DNA damage response in oxidative stress-induced vascular endothelial cell senescence. J Biol Chem 2010; 285: 29662-70. doi: 10.1074/jbc.M110.125138
- 44. Spyridopoulos I, Isner JM, Losordo DW. Oncogenic ras induces premature senescence in endothelial cells: role of p21(Cip1/Waf1). Basic Res Cardiol 2002; 97: 117-24. doi: 10.1007/s003950200001
- 45. Dai DF, Rabinovitch PS, Ungvari Z. Mitochondria and cardiovascular aging. Circ Res 2012; 110: 1109-24. doi: 10.1161/circresaha.111.246140
- 46. Jia G, Aroor AR, DeMarco VG, Martinez-Lemus LA, Meininger GA, Sowers JR. Vascular stiffness in insulin resistance and obesity. Front Physiol 2015; 6: 231. doi: 10.3389/fphys.2015.00231
- 47. Donato AJ, Morgan RG, Walker AE, Lesniewski LA. Cellular and molecular biology of aging endothelial cells. J Mol Cell Cardiol 2015; 89: 122-35. doi: 10.1016/j.yjmcc.2015.01.021
- 48. Vasan RS, Beiser A, Seshadri S, Larson MG, Kannel WB, D'Agostino RB, et al. Residual lifetime risk for developing hypertension in middle-aged women and men: The Framingham Heart Study. JAMA 2002; 287: 1003-10. doi: 10.1001/jama.287.8.1003
- 49. Zhou J, Wang Q, Ding Y, Zou MH. Hypochlorous acid via peroxynitrite activates protein kinase $C\theta$ and insulin resistance in adipocytes. J Mol Endocrinol 2015; 54: 25-37. doi: 10.1530/jme-14-0213.
- 50. Jia G, Habibi J, Aroor AR, Hill MA, DeMarco VG, Lee LE, et al. Enhanced endothelium epithelial sodium channel signaling prompts left ventricular diastolic dysfunction in obese female mice. Metabolism: Clin Exper 2018; 78: 69-79. doi: 10.1016/j.metabol.2017.08.008
- 51. Briley-Saebo KC, Shaw PX, Mulder WJ, Choi SH, Vucic E, Aguinaldo JG, et al. Targeted molecular probes for imaging atherosclerotic lesions with magnetic resonance using antibodies that recognize oxidation-specific epitopes. Circulation 2008; 117: 3206-15. doi: 10.1161/circulationaha.107.757120

- 52. Bochkov VN. Inflammatory profile of oxidized phospholipids. Thromb Hemost 2007; 97: 348-54.
- 53. Pal PB, Sonowal H, Shukla K, Srivastava SK, Ramana KV. Aldose reductase mediates NLRP3 inflammasome-initiated innate immune response in hyperglycemia-induced Thp1 monocytes and male mice. Endocrinol 2017; 158: 3661-75. doi: 10.1210/en.2017-00294
- 54. Khan SY, Awad EM, Oszwald A, Mayr M, Yin X, Waltenberger B, et al. Premature senescence of endothelial cells upon chronic exposure to TNFα can be prevented by N-acetyl cysteine and plumericin. Sci Rep 2017; 7: 39501. doi: 10.1038/srep39501
- Poznyak AV, Nikiforov NG, Markin AM, Kashirskikh DA, Myasoedova VA, Gerasimova EV, et al. Overview of oxLDL and its impact on cardiovascular health: Focus on atherosclerosis. Front Pharmacol 2020; 11:613780. doi: 10.3389/fphar.2020.613780
- 56. Kovanen PT. Mast cells as potential accelerators of human atherosclerosis-from early to late lesions. Int J Mol Sci 2019; 20. doi: 10.3390/ijms20184479
- 57. Canepa M, Viazzi F, Strait JB, Ameri P, Pontremoli R, Brunelli C, et al. Longitudinal association between serum uric acid and arterial stiffness: results from the baltimore longitudinal study of aging. Hypertension 2017; 69: 228-35. doi: 10.1161/hypertensionaha.116.08114
- 58. Aroor AR, Jia G, Habibi J, Sun Z, Ramirez-Perez FI, Brady B, et al. Uric acid promotes vascular stiffness, maladaptive inflammatory responses and proteinuria in western diet fed mice. Metabolism: Clin Exp 2017; 74: 32-40. doi: 10.1016/j.metabol.2017.06.006
- 59. Neogi T, Ellison RC, Hunt S, Terkeltaub R, Felson DT, Zhang Y. Serum uric acid is associated with carotid plaques: the National Heart, Lung, and Blood Institute Family Heart Study. J Rheumatol 2009; 36: 378-84. doi: 10.3899/jrheum.080646
- 60. Han MS, Jung DY, Morel C, Lakhani SA, Kim JK, Flavell RA, et al. JNK expression by macrophages promotes obesity-induced insulin resistance and inflammation. Science 2013; 339: 218-22. doi: 10.1126/science.1227568
- 61. Bolton E, Rajkumar C. The ageing cardiovascular system. Rev Clin Gerontol 2011; 21: 99-109. doi: 10.1017/ S0959259810000389
- 62. Lund G, Zaina S. Atherosclerosis: an epigenetic balancing act that goes wrong. Curr Atheroscl Rep 2011; 13: 208-14. doi: 10.1007/s11883-011-0174-3
- 63. Beekman M, Blanché H, Perola M, Hervonen A, Bezrukov V, Sikora E, et al. Genome-wide linkage analysis for human longevity: Genetics of Healthy Aging Study. Aging cell 2013; 12: 184-93.
- 64. Kurz DJ, Kloeckener-Gruissem B, Akhmedov A, Eberli FR, Bühler I, Berger W, et al. Degenerative aortic valve stenosis,

- but not coronary disease, is associated with shorter telomere length in the elderly. Arterioscler Thromb Vasc Biol 2006; 26: e114-e7.
- 65. Bianchessi V, Badi I, Bertolotti M, Nigro P, D'Alessandra Y, Capogrossi MC, et al. The mitochondrial IncRNA ASncmtRNA-2 is induced in aging and replicative senescence in endothelial cells. J Mol Cell Cardiol 2015; 81: 62-70.
- 66. Furukawa S, Fujita T, Shimabukuro M, Iwaki M, Yamada Y, Nakajima Y, et al. Increased oxidative stress in obesity and its impact on metabolic syndrome. J Clin Invest 2017; 114: 1752-61.
- 67. Chisolm GM, Steinberg D. The oxidative modification hypothesis of atherogenesis: an overview. Free Rad Biol Med 2000; 28: 1815-26.
- 68. Matsuda M, Shimomura I. Increased oxidative stress in obesity: implications for metabolic syndrome, diabetes, hypertension, dyslipidemia, atherosclerosis, and cancer. Obes Res Clin Pract 2013; 7: e330-e41.
- 69. Nanda N, Bobby Z, Hamide A, Koner BC, Sridhar MG. Association between oxidative stress and coronary lipid risk factors in hypothyroid women is independent of body mass index. Metabol Clin Exp 2007; 56: 1350-5.
- 70. Mattisson IY, Christoffersen C. Apolipoprotein M and its impact on endothelial dysfunction and inflammation in the cardiovascular system. Atheroscler 2021; 334: 76-84.
- 71. Silver AE, Beske SD, Christou DD, Donato AJ, Moreau KL, Eskurza I, et al. Overweight and obese humans demonstrate increased vascular endothelial NAD (P) H oxidase-p47phox expression and evidence of endothelial oxidative stress. Circulation 2007; 115: 627-37.
- 72. López-Domènech S, Martínez-Herrera M, Abad-Jiménez Z, Morillas C, Escribano-López I, Díaz-Morales N, et al. Dietary weight loss intervention improves subclinical atherosclerosis and oxidative stress markers in leukocytes of obese humans. Int J Obes 2019; 43: 2200-9.
- 73. Amizuka N, Hasegawa T, Oda K, de Freitas PHL, Hoshi K, Li M, et al. Histology of epiphyseal cartilage calcification and endochondral ossification. Front Bioscience-Elite 2012; 4: 2085-100.
- 74. Shi X, Gao J, Lv Q, Cai H, Wang F, Ye R, et al. Calcification in atherosclerotic plaque vulnerability: Friend or foe? Front Physiol 2020; 11: 56. doi: 10.3389/fphys.2020.00056
- 75. Bagyura Z, Kiss L, Lux Á, Csobay-Novák C, Jermendy Á L, Polgár L, et al. Association between coronary atherosclerosis and visceral adiposity index. Nutr Metabol Cardiovasc Dis 2020; 30: 796-803. doi: 10.1016/j. numecd.2020.01.013
- 76. Sommer F, Bäckhed F. The gut microbiota—masters of host development and physiology. Nat Rev Microbiol 2013; 11: 227-38.

- 77. Turnbaugh PJ, Hamady M, Yatsunenko T, Cantarel BL, Duncan A, Ley RE, et al. A core gut microbiome in obese and lean twins. Nature 2009; 457: 480-4. doi: 10.1038/nature07540
- Sokol H, Pigneur B, Watterlot L, Lakhdari O, Bermúdez-Humarán LG, Gratadoux JJ, et al. Faecalibacterium prausnitzii is an anti-inflammatory commensal bacterium identified by gut microbiota analysis of Crohn disease patients. Proc Nat Acad Sci USA 2008; 105: 16731-6. doi: 10.1073/pnas.0804812105
- 79. Karlsson FH, Tremaroli V, Nookaew I, Bergström G, Behre CJ, Fagerberg B, et al. Gut metagenome in European women with normal, impaired and diabetic glucose control. Nature 2013; 498: 99-103. doi: 10.1038/nature12198 80.Wang Z, Klipfell E, Bennett BJ, Koeth R, Levison BS, DuGar B, et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. Nature 2011; 472: 57-63.
- 81. Tang WW, Hazen SL. The contributory role of gut microbiota in cardiovascular disease. J Clin Invest 2014; 124: 4204-11.
- 82. Tang WW, Wang Z, Levison BS, Koeth RA, Britt EB, Fu X, et al. Intestinal microbial metabolism of phosphatidylcholine and cardiovascular risk. New Engl J Med 2013; 368: 1575-84.
- 83. Aguilar EC, Leonel AJ, Teixeira LG, Silva AR, Silva JF, Pelaez JM, et al. Butyrate impairs atherogenesis by reducing plaque inflammation and vulnerability and decreasing NFkB activation. Nutr Metabol Cardiovasc Dis 2014; 24: 606-13. doi: 10.1016/j.numecd.2014.01.002
- 84. Aguilar EC, Santos LC, Leonel AJ, de Oliveira JS, Santos EA, Navia-Pelaez JM, et al. Oral butyrate reduces oxidative stress in atherosclerotic lesion sites by a mechanism involving NADPH oxidase down-regulation in endothelial cells. J Nutr Biochem 2016; 34: 99-105. doi: 10.1016/j. jnutbio.2016.05.002
- 85. Seldin MM, Meng Y, Qi H, Zhu W, Wang Z, Hazen SL, et al. Trimethylamine N-oxide promotes vascular inflammation through signaling of mitogen-activated protein kinase and nuclear factor-κB. J Am Heart Assoc 2016; 5. doi: 10.1161/jaha.115.002767
- 86. Murphy EA. Some difficulties in the investigation of genetic factors in coronary artery disease. Can Med Assoc J 1967; 97: 1181-92.
- 87.Libby P, Egan D, Skarlatos S. Roles of infectious agents in atherosclerosis and restenosis: an assessment of the evidence and need for future research. Circulation 1997; 96: 4095-103. doi: 10.1161/01.cir.96.11.4095
- 88. Ulker S, McKeown PP, Bayraktutan U. Vitamins reverse endothelial dysfunction through regulation of eNOS and NAD(P)H oxidase activities. Hypertension 2003; 41: 534-9. doi: 10.1161/01.hyp.0000057421.28533.37

- 89. Wray DW, Nishiyama SK, Harris RA, Zhao J, McDaniel J, Fjeldstad AS, et al. Acute reversal of endothelial dysfunction in the elderly after antioxidant consumption. Hypertension 2012; 59: 818-24. doi: 10.1161/hypertensionaha.111.189456
- 90. Su JB. Vascular endothelial dysfunction and pharmacological treatment. World J Cardiol 2015; 7: 719-41. doi: 10.4330/wjc.v7.i11.719
- 91. Cao F, Wu K, Zhu YZ, Bao ZW. Roles and mechanisms of dipeptidyl peptidase 4 inhibitors in vascular aging. Front Endocrinol 2021; 12: 731273. doi: 10.3389/fendo.2021.731273
- 92. Janaszak-Jasiecka A, Płoska A, Wierońska JM, Dobrucki LW, Kalinowski L. Endothelial dysfunction due to eNOS uncoupling: molecular mechanisms as potential therapeutic targets. Cell Mol Biol Lett 2023; 28: 21. doi: 10.1186/s11658-023-00423-2
- 93. Takase S, Matoba T, Nakashiro S, Mukai Y, Inoue S, Oi K, et al. Ezetimibe in combination with statins ameliorates endothelial dysfunction in coronary arteries after stenting: The CuVIC Trial (Effect of Cholesterol Absorption

- Inhibitor Usage on Target Vessel Dysfunction After Coronary Stenting), a multicenter randomized controlled trial. Arterioscler Thromb Vasc Biol 2017;37(2):350-8. doi: 10.1161/atvbaha.116.308388
- 94. Patel AA, Budoff MJ. Effects of eicosapentaenoic acid and docosahexaenoic acid on lipoproteins in hypertriglyceridemia. Curr Opin Endocrinol Diabet Obes 2016; 23: 145-9. doi: 10.1097/med.00000000000000233
- 95. Nelson JR, Wani O, May HT, Budoff M. Potential benefits of eicosapentaenoic acid on atherosclerotic plaques. Vasc Pharmacol 2017; 91: 1-9. doi: 10.1016/j.vph.2017.02.004
- 96. Borow KM, Nelson JR, Mason RP. Biologic plausibility, cellular effects, and molecular mechanisms of eicosapentaenoic acid (EPA) in atherosclerosis. Atheroscler 2015; 242: 357-66. doi: 10.1016/j.atherosclerosis.2015.07.035